

Patent



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DATE

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicants : Barbara Grimpe et al.
Serial No. : 10/698,190
Filing Date : October 31, 2003
For : METHODS OF INHIBITING GLIAL SCAR FORMATION
Group Art Unit : 1633
Examiner : Scott Long
Attorney Docket No. : CWR-7779NP

Mail Stop Amendment
Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

DECLARATION UNDER 37 CFR §1.132

Sir:

We Barbara Grimpe and Jerry Silver declare as follows:

1. We are the inventors and Applicants of the invention entitled "METHOD OF INHIBITING GLIAL SCAR FORMATION", disclosed and claimed in U.S. Application Serial No. 10/698,190, filed October 31, 2003.
2. The invention described and claimed in said application claims priority from U.S. Provisional Application Serial No. 60/423,082, filed November 1, 2002, and U.S. Provisional Application Serial No. 60/471,447, filed May 16, 2003.

BEST AVAILABLE COPY

3. We are the inventors of the subject matter disclosed in *The Journal of Neuroscience*, April 15, 2002, 22(8):3144-3160, of which we are listed as authors, and which was received for publication on September 24, 2001.

We hereby declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under §1001 of Title 18 of the United States Code, and that such willful or false statements may jeopardize the validity of the application or any patent issued thereon.

Barbara Grimpe
Barbara Grimpe

Barbara Grimpe

Jerry Silver

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3. Prior to April 11, 2002, we had reduced to practice a method of reducing GAG content in a glial scar comprising inhibiting one or more of the following: inhibiting the expression of primary proteoglycans; inhibiting the expression and/or activity of a chain initiation enzyme; and inhibiting the expression and/or activity of a chain elongation enzyme.

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B. Grimpe
Barbara Grimpe

Jerry Silver

1/16/07

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Jane Grimpe January 16, 2007
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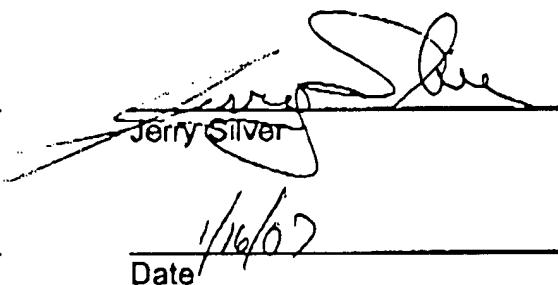
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Jerry Silver
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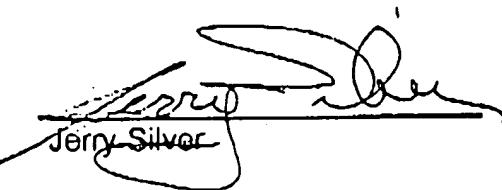
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Date


Jerry Silver
1/16/07

The Critical Role of Basement Membrane-Independent Laminin $\gamma 1$ Chain during Axon Regeneration in the CNS

Barbara Grimpe,¹ Sucai Dong,³ Catherine Doller,¹ Katherine Temple,² Alfred T. Malouf,² and Jerry Silver¹

Departments of ¹Neurosciences and ²Pediatrics, School of Medicine, Case Western Reserve University, Cleveland, Ohio 44106, and ³Department of Neurobiology, University of Pittsburgh, Pittsburgh, Pennsylvania 15261

We have addressed the question of whether a family of axon growth-promoting molecules known as the laminins may play a role during axon regeneration in the CNS. A narrow sickle-shaped region containing a basal lamina-independent form of laminin exists in and around the cell bodies and proximal portion of the apical dendrites of CA3 pyramidal neurons of the postnatal hippocampus. To understand the possible function of laminin in axon regeneration within this pathway, we have manipulated laminin synthesis at the mRNA level in a slice culture model of the lesioned mossy system. In this model early postnatal mossy fibers severed near the hilus can regenerate across the lesion and elongate rapidly within strata lucidum and pyramidale. In slice cultures of the postnatal day 4 hippocampus, 2 d before lesion and then continuing for 1–5 d after lesion, translation of the $\gamma 1$ chain product of laminin was reduced by

using antisense oligodeoxyribonucleotides and DNA enzymes. In the setting of the lesioned organotypic hippocampal slice, astroglial repair of the lesion and overall glial patterning were unperturbed by the antisense or DNA enzyme treatments. However, unlike controls, in the treated, lesioned slices the vast majority of regenerating mossy fibers could not cross the lesion site; those that did were very much shorter than usual, and they took a meandering course. In a recovery experiment in which the DNA enzyme or antisense oligos were washed away, laminin immunoreactivity returned and mossy fiber regeneration resumed. These results demonstrate the critical role of laminin(s) in an axon regeneration model of the CNS.

Key words: extracellular matrix; hippocampus; organotypic slice cultures; reactive astrocytes; glial scar; dendritic spines; axon guidance; antisense ODN; DNA enzyme

The environment surrounding CNS lesions is composed of a complex mixture of both axon growth-promoting and inhibitory cell surface molecules (Eddleston and Mucke, 1993; Fouad et al., 2001; Tang et al., 2001) as well as extracellular matrix (ECM) proteins (Rudge and Silver, 1990; Pindzola et al., 1993; Stichel and Muller, 1994; McKeon et al., 1995; Canning et al., 1996; Giger et al., 1998; Fitch et al., 1999; Niederost et al., 1999; Pasterkamp et al., 1999; Asher et al., 2000; Lemons et al., 2001) for which the functional interactions with the cut end of the axon are dependent on the particular region that is damaged, the maturational state of a given axon tract, as well as the proximity of the reactive cells to the heart of the lesion. Although robust axon regeneration through or around lesions usually fails after CNS injury, there are interesting exceptions (Kalil and Reh, 1979; Kunkel-Bagden et al., 1992; Dow et al., 1994; Chauvet et al., 1995; Doucette, 1995; Davies et al., 1997, 1999; Fry and Saunders, 2000; Magavi et al., 2000; Scharff et al., 2000; Fischer et al., 2001). What might be the molecular mechanisms that underlie the dramatic axon growth-promoting potential of certain forms of the regeneration-permissive reactive tract phenotype that develop, on occasion, in the CNS? The laminins are a 15 member family of

large cruciform glycoproteins (Timpl et al., 1979; Burgeson et al., 1994; Koch et al., 1999) that, in the nervous system, are produced mainly by Schwann cells (Cornbrooks et al., 1983), olfactory ensheathing glia (Obremski and Bunge, 1995; Ramon-Cueto and Avila, 1998; Li et al., 1999), astroglia (Liesi and Silver, 1988; Liesi and Risteli, 1989; Liesi, 1990), and neurons (Zhou, 1990; Jucker et al., 1991; Hagg et al., 1997). The potent axon growth and guidance capacities of the various laminin subtypes have been demonstrated repeatedly in tissue culture assays (Edgar et al., 1984; Gundersen, 1987; Smith et al., 1990; Liesi et al., 1992; Calof et al., 1994; Kennedy and Tessier-Lavigne, 1995; Kuhn et al., 1995; Matsuzawa et al., 1996; Ivins et al., 1998; Powell et al., 2000). *In vivo*, although there is a great deal of evidence in support of the critical role of laminin in PNS regeneration (Zhou, 1990; Jucker et al., 1991; Patton et al., 1997; Ferguson and Muir, 2000) (see also Bonner and O'Connor, 2001), relatively little is known about the possible role of the laminins in the process of axon regeneration in the mammalian CNS that can occur within the context of a nonscarred, intratract glial framework.

One model for investigating the influence of laminin during successful regeneration of the cut axon in the brain is the mossy fiber pathway in the hippocampus. Mossy fibers originate from granule cells in the dentate gyrus and synapse in the hilus and in the stratum lucidum of the CA3 area, stopping abruptly at the CA3–CA2 border (Gaarskjaer, 1986). Importantly, the mossy fiber pathway shows a remarkable degree of plasticity throughout life (Steward, 1976; Amaral and Dent, 1981; Laurberg and Zimmer, 1981; McWilliams and Lynch, 1983; Eriksson et al., 1998) and is capable of complete regeneration of its normal synaptic pattern when transected in organotypic slice cultures (Zimmer and Gähwiler, 1987; Nguyen et al., 1996). Furthermore, the path-

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way over which the axons regenerate is decorated richly with a form of intraparenchymal laminin that is one of the few in the CNS that is clearly and easily demonstrable with immunohistochemistry (Zhou, 1990; Hagg et al., 1997; Nakagami et al., 2000). Characterization of a regeneration-stimulating role for particular ECM molecules is important if we are to understand how the CNS may be able to repair itself intrinsically or be induced to repair itself after devastating injuries.

To investigate the possible role of laminin in this particular case of CNS axon regeneration, we found the specific reduction of one protein or one family of proteins in postnatal animals to be of high interest. Gene-targeting experiments for different laminin chains such as LAMC1, which codes for the laminin $\gamma 1$ chain, result in the death of the embryo (Smyth et al., 1999). Although these observations highlight the absolute biological prerequisite for functional laminin interactions during development, the demise of the laminin-deficient embryo limits the possibility for further investigations of laminin function.

The antisense-targeting technology in combination with DNA enzymes allows for the large-scale downregulation or even complete inhibition of particular proteins *in vivo* or *in situ*. In our experiments, synthetic oligodeoxyribonucleotides (ODNs) were used for specific hybridization with an expressed target, the laminin $\gamma 1$ chain mRNA. Antisense ODNs or DNA enzymes physically bind to the target gene transcript, producing a nucleic acid duplex and, in the latter case, actually cut the target mRNA specifically, which leads to the inhibition of translation (Probst and Skutella, 1996; Grimpe et al., 1999). For the present experiments we have used these techniques to interrupt the formation of nearly all known forms of laminin during regeneration of the mossy fibers in an organotypic hippocampal slice culture model. With the use of this strategy we demonstrate experimentally, for the first time, the critical role of laminin(s) in regeneration of an axon tract in the mammalian CNS.

MATERIALS AND METHODS

Hippocampal slice cultures and immunohistochemistry. Hippocampal slice cultures were prepared from 4-d-old Sprague Dawley rat pups by the method of Stoppini et al. (1991). Rats were decapitated, and 400 μ m transverse hippocampal slices were prepared with a Siskiyou Designs brain slicer (Grants Pass, OR). Slices were placed on 25 mm Nunc Anopore tissue culture inserts (Naperville, IL) in six-well tissue culture plates and fed every 3 d with 1.5 ml of growth medium [50% MEM, 25% HBSS (Invitrogen, San Diego, CA), and 25% horse serum]. For staining the mossy fibers, 0.1–0.5 μ l of Micro Ruby 3 kDa (Molecular Probes, Eugene, OR) coupled to dextran was injected with the use of a Marzhauser micromanipulator into the granule cell layer by following the general procedure described by Harrigan et al. (1995). The cultures were incubated for 1.5 hr in an incubator at 5% CO₂ and 37°C, followed by fixation in 4% paraformaldehyde (PFA) in 0.1 M PB (1.9 mM NaH₂PO₄ \times H₂O and 8 mM Na₂HPO₄ \times 7 H₂O) for 3–4 hr. The slice cultures were washed two times in 10% sucrose, once in 30% sucrose, and then incubated in 30% sucrose overnight. To enhance antibody penetration, we then freeze-thawed the cultures three times in 30% sucrose on dry ice, washed them two times in 10% sucrose, once in 0.1 M TB (80 mM Trizma-HCl, 20 mM Trizma Base), and once in 0.1 M TBS [containing (in mM) 80 Trizma-HCl, 20 Trizma Base, 150 NaCl, 2 KCl] for 15 min. After the cultures were blocked for 1 hr with blocking solution (3% goat serum, 3% BSA, 1% Triton X-100, 1% DMSO in 0.05 M TBS), the slices were washed again in TBS for 1 hr with a change of the solution every 5 min (Kunkel et al., 1994). Then the slice cultures were incubated with the first antibody against mouse glial fibrillary acidic protein (GFAP; 1:500, Sigma, St. Louis, MO) for 2 or 3 d. This was followed by rinsing in TBS for 1 hr and then by overnight incubation with the second antibody (anti-mouse) coupled with Oregon green or Alexa 488 (1:500, Sigma; 1:500, Molecular Probes). This second antibody was preincubated with rat serum (Sigma) for 30 min at room temperature

before use. Finally, the cultures were washed in TBS for 2 hr and mounted with ImmunoFluor Mounting Medium (ICN Biochemicals, Aurora, OH). The double immunofluorescence labeling was analyzed by confocal laser-scanning microscopy with a Zeiss (Oberkochen, Germany) microscope in an inverted configuration. The z-section rotation was done with Zeiss software.

Laminin staining of hippocampal sections and slice cultures. Hippocampal sections (400 μ m) of postnatal day 4 (P4) as well as adult (6 month to 1 year) Sprague Dawley rats or living P4 slices (400 μ m) cultured for 1–7 d were used unfixed, fresh-frozen, or fixed with 2% PFA and 0.05% glutaraldehyde for ~1 hr, followed by rinsing in 1× PBS [containing (in mM) 1 K₂HPO₄, 10 NaHPO₄, and 2.7 KCl, plus 0.137 M NaCl, pH 7.4]. The unfixed fresh-frozen P4 hippocampi were sectioned with a cryostat (Bright Instrument, UK; Hacker Instruments, Fairfield, NJ), followed by the staining procedure described below. The fixed cultured slices were removed gently from the insert with a brush and washed for 3–4 hr or overnight with 1× PBS on an orbital shaker. Then the slice cultures or sections were blocked with 10% normal goat serum in 1× PBS for at least 1 hr at room temperature on a shaker. The primary antibody against human laminin $\gamma 1$ chain made in mouse, clone 2E8 (1:100, Invitrogen), Engelbreth–Holm–Swarm (EHS) laminin made in rabbit (1:25, Sigma), neurocan made in rabbit [NC 2; 1:2000, a generous gift by Uwe Rauch (Oleszewski et al., 1999)], or rabbit serum as a negative control (in a dilution of the respective antibodies, 1:100 and 1:2000) as well as only the second antibody, which never showed any staining, was left on the sections or slices for 2 d at 4°C on an orbital shaker. After 2–3 hr of rinsing in 1× PBS (~6–8 changes), the tissues were put in secondary antibody coupled with Alexa 594 (1:200, goat against mouse or rabbit IgG; Molecular Probes) overnight at 4°C on a shaker. The next day the tissues were again rinsed for several hours and then mounted with Citifluor mounting medium (Ted Pella, Redding, CA). The 2E8 antibody has been characterized by Engvall et al. (1986), and electron microscopic (EM) rotary shadow images showed the binding domain of this $\gamma 1$ antibody to be in the P1 fragment of the laminin molecule.

Antisense and DNA enzyme design and treatment. Antisense oligonucleotides and DNA enzymes were designed as end-capped phosphorothioate ODNs of three or two bases on each end. They were obtained from a commercial supplier (MWG Biotech, Ebersberg, Germany). The sequence corresponded to the 3' end of the published laminin $\gamma 1$ chain mRNA sequence (accession number X94551; Vanden Heuvel et al., 1996). It is well away from the coding sequence of all netrins (Manitt et al., 2001) and had no homology to other mammalian sequences registered in the GenBank databases of National Institutes of Health (Altenschul et al., 1997).

Hippocampal slice cultures were pretreated for 2 d with the specific concentration (0.05, 0.1, 0.5, 2, and 8 μ M) of antisense [Lamy1a2, 5'-ATG GTC CGG TTG ATG GCG GG-3', nucleotides position in the sequence 401–420 (Vanden Heuvel et al., 1996)] or mixed base (Lamy1mb2, 5'-GCG CCC ATC AAC CGG AGT CA-3') ODNs every day or, in the untreated control slices, with nothing. The respective DNA enzyme targeted the same sequence of the laminin $\gamma 1$ chain mRNA. However, for the catalytic digestion of the targeted mRNA the DNA enzyme contained a loop structure (Santoro and Joyce, 1997) and had the following sequence of DNA enzyme: 5'-GGT CCG GTG GCT AGC TAC AAC GAG ATG GCG G-3', nucleotides position in the sequence 402–418 (Vanden Heuvel et al., 1996). The control DNA mixed base enzyme lacked the ability to digest target mRNA and the inability to bind to any other sequence [control DNA enzyme (mixed base), 5'-TCG ACG GTA GCA ACA TCG ATC GGG ATG TGA C-3']. Lesions were made in the hilar portion of the CA3 region of slice cultures with a surgical blade (stainless steel, size 15, Feather, Fisher Scientific, Pittsburgh, PA). The cultures were treated every day for 1 or 5 d further with 2 μ l of the specific concentration of antisense, DNA enzyme, or mixed base ODNs or were allowed to develop untreated. The slice cultures either were subjected to immunocytochemistry or were used for RNA isolation.

Quantification. To characterize the density of mossy fibers that cross lesion sites in contrast to the density of fibers that do not, we lesioned slice cultures after 2 d of pretreatment. The slices were treated further with antisense or mixed base ODNs for 5 d more, and the mossy fibers were stained as described above. Photomicrographs were taken with a Nikon Optiphot-2 fluorescence microscope connected to a SPOT color charge-coupled device camera (Diagnostic Instruments, Sterling Heights, MI).

For the semiquantitative analysis of axon regeneration, an area on both sides of the lesion and a background region were defined, and the

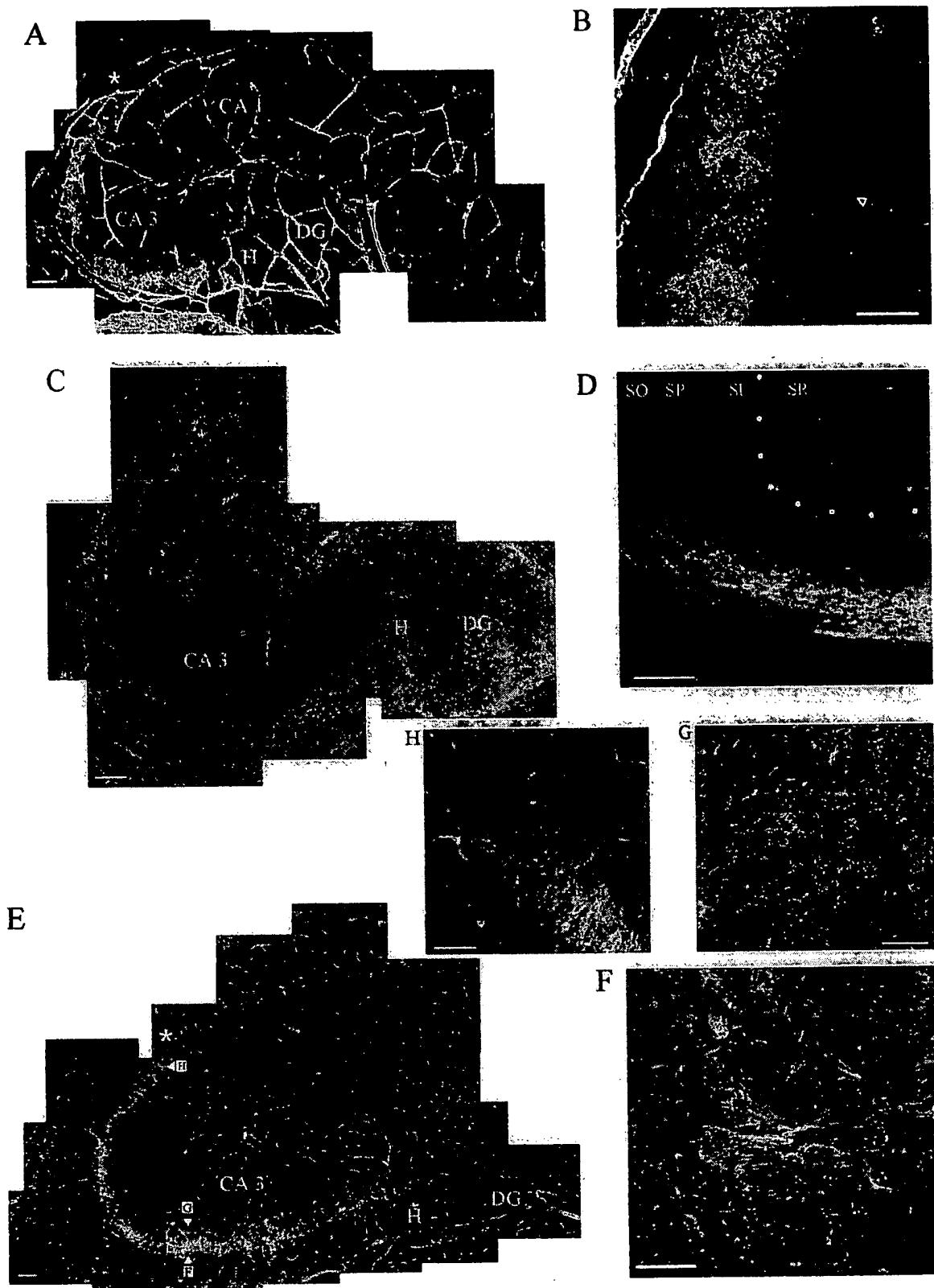


Figure 1. Shown are freshly fixed P4 (*A–D*) and adult (*E–H*) hippocampal sections (20- μ m-thick) stained for the $\gamma 1$ chain of laminin and for neurocan and imaged with a confocal laser microscope. *A*, Pathway of punctate laminin staining in the stratum pyramidale and stratum lucidum, starting in the hilar region (*H'*) of the dentate gyrus (*DG*) and ending at the CA3–CA2 border (asterisk). The blood vessels of the hippocampus are stained also. Scale bar, 100 μ m. *B*, Higher magnification of this pathway, which shows the cell bodies and proximal portion of the apical dendrites of the pyramidal neurons stained for laminin $\gamma 1$ (arrowhead). Scale bar, 25 μ m. *C*, A P4 hippocampal section is stained for neurocan (green) and laminin (Figure legend continues.)

intensities were measured with TINA 2.09 (Raytest GmbH, Straubenhardt, Germany). In the profile window the baseline was subtracted from the intensity measurement. For the statistical evaluation (percentage, average, SD) Excel (Microsoft, Redmond, WA) was used. The percentages of regeneration at three specific distances one point proximal (point A) and two points distal to the lesion (points B and C) were calculated for each slice culture. A schematic diagram, which explains where the measurement points were taken, is presented in Figure 6. For the average amount of regeneration at each distance, the individual percentage rates from each slice culture from each concentration group of antisense or mixed base ODN were totaled and divided by the numbers of slices in their group. These results are shown in Table 1 also.

Immunoprecipitation. Twelve hippocampal slices in culture for 7 d were treated with 0.05 or 8 μM DNA enzyme and control DNA enzyme (mixed base). The slices were collected in immunoprecipitation buffer [containing (in mM) 10 Tris-HCl, pH 7.6, 150 NaCl, 0.2 PMSF, 1 EDTA, pH 8, plus 1% Triton X-100, 0.5% NP-40] and kept at –80°C until they were used. Then they were triturated, and the protein concentration was measured with the BCA assay (Pierce, Rockford, IL) on a spectrophotometer at a wavelength of 590 nm. Identical amounts of protein from DNA enzyme-treated and control DNA enzyme-treated (mixed base) slices were extracted with 500 μl of 0.5 M NaCl and 0.05 M Tris, pH 7.7, solution. After centrifugation the supernatant was dialyzed against 1× PBS overnight. On the next day the proteins were biotinylated with 200 μl of EZ-linked sulfo-N-hydroxysuccinimide-biotin (Pierce) for 30 min at room temperature on a shaker. The solution was cleared with a 3 hr incubation with an actin IgG2a antibody and protein A-Sepharose at 4°C on a shaker, followed by a short centrifugation. The supernatant was incubated for 1 hr with the γ1 chain laminin antibody (clone 2E8; 2 μl of mouse ascites, Invitrogen) at 4°C on a shaker. The solution was incubated with protein A-Sepharose overnight at 4°C on a shaker. The next day each solution was washed two times, with a 20 sec centrifugation in between, with the three following solutions: wash buffer 1 (150 mM NaCl, 50 mM Tris-HCl, pH 7.4, 1% NP-40), wash buffer 2 (500 mM NaCl, 50 mM Tris-HCl, pH 7.4, 0.1% NP-40), and wash buffer 3 (50 mM Tris-HCl, pH 7.4, 0.1% NP-40). After removal of the third wash buffer the precipitates were incubated for 5 min at 95°C in Laemmli buffer (0.1 M dithiothreitol; Laemmli, 1970) under reducing conditions. The probes were separated electrophoretically in a 4–12% SDS-PAGE (Bio-Rad, Hercules, CA) in 1× TGS running buffer (3.03 gm of Tris Base, 14.4 gm of glycine, 1 gm of SDS for 1 l), starting with 60 V and increasing to 100 V. The gel was blotted electrophoretically on a nitrocellulose membrane (Bio-Rad) in 10 mM sodium borate (borax) buffer overnight at 4°C. On the next day the membrane was blocked with 3% BSA in TBS (10 mM Tris, pH 8, 150 mM NaCl) for 2 hr at room temperature, followed by a 2 hr incubation of 250 mM streptavidin-coupled horseradish peroxidase in 1% BSA in TBS-T (10 mM Tris, pH 8, 150 mM NaCl, 0.1% Tween 20) for 2 hr further on a shaker. To remove unbound streptavidin–horseradish peroxidase, we rinsed the membrane in TBS-T (~6–8 changes), followed by developing with the ECL kit (Amersham Biosciences, Arlington Heights, IL).

Western blot analysis. Ten microliters of the supernatant from the immunoprecipitated 0.05 μM treated DNA enzyme and control DNA enzyme [DNA mixed base (mb)] protein extract were mixed with 10 μl of Laemmli buffer (Laemmli, 1970) and incubated for 5 min at 95°C. The solution was loaded on a 4–12% SDS-PAGE (Bio-Rad) in 1× TGS running buffer, starting with 60 V and increasing to 100 V. The gel either was stained with Coomassie blue R-250 in 9.5% acidic acid and 43% methanol, followed by a destaining solution of 10% acidic acid and 10% methanol, or was blotted electrophoretically on a nitrocellulose membrane (Bio-Rad) in 10 mM sodium borate (borax) buffer overnight at 4°C. On the next day the membrane was blocked with 5% low fat milk powder in TBS-T buffer for at least 2 hr at room temperature. Then the mem-

brane was incubated with a mouse monoclonal antibody against GFAP (1:500, Chemicon, Temecula, CA), β-actin (1:500, Sigma), in 5% low fat milk powder in TBS-T overnight on a shaker at 4°C. On the next day the membrane was washed with TBS-T for 5 hr (several changes) and incubated with the second goat antibody (which recognizes mouse IgG labeled with horseradish peroxidase; 1:500, Chemicon) in 5% low fat milk powder in TBS-T overnight at 4°C. To remove unbound secondary antibody, we rinsed the membrane for 5 hr in TBS-T (several changes), followed by incubation of the membrane with the ECL kit (Amersham Biosciences).

In situ hybridization. The probe for nonradioactive *in situ* hybridization against the γ1 chain mRNA was a gift from Yoshihiko Yamada (Sasaki and Yamada, 1987). It contains the mouse V and VI domains of the γ1 chain mRNA of laminin (position 333–1668) cloned in pBluescript II SK (Stratagene, La Jolla, CA). After a Qiagen plasmid preparation (Chatsworth, CA) in accordance with the manufacturer's protocol, the plasmid was linearized by a *Nor*I digestion for the antisense probe and with *Xba*I for the sense control probe. To purify the reaction product, we used a purification kit from Stratagene. This was followed by the labeling reaction with digoxigenin (DIG; Boehringer Mannheim, Indianapolis, IN) for 2 hr at 37°C to a cRNA probe by using T7 polymerase. The RNA was precipitated by using Pellet Paint (Novagen, Madison, WI), 0.1 volume of 3 M sodium acetate, pH 5.2, and 2 volumes of cold 100% ethanol. For quality control the probe was separated electrophoretically on an agarose gel.

Freshly prepared P4 mouse hippocampi (C57BL/6J) were fixed for 12 hr in 4% PFA/PBS, followed by incubation in 30% sucrose in 1× PBS overnight. On the next day 25 μm coronal sections were cut on a cryostat (Bright Instrument, UK; Hacker Instruments), transferred to cold Superfrost plus slides (Fisher brand), and stored at –80°C. All solutions were made with DEPC-treated (diethyl pyrocarbonate) water, followed by autoclaving. The sections were air dried for 1 hr at room temperature and post-fixed in 4% PFA/PBS for 10 min at room temperature. After being rinsed in PBS three times, the sections were digested with proteinase K (1 μg/ml in 50 mM Tris, pH 7.5, and 5 mM EDTA) for 7 min, followed by a further fixing step in 4% PFA/PBS for 5 min at room temperature and then three more rinses in PBS for 3 min. The acetylation reaction was performed by mixing 3.3 ml of triethanolamine with 0.513 ml of a 10% HCl in 246 ml of DEPC-treated water. Then 0.75 ml of acetic anhydride was added and mixed by dipping the slides carefully several times, followed by a 10 min incubation at room temperature. The slides were covered with prehybridization solution (50% formamide, 5× SSC, 5× Denhardt's reagent, 250 μg/ml baker's yeast RNA, and 500 μg/ml salmon sperm) and incubated for 2 hr at room temperature in a humidified chamber. This prehybridization solution was poured off and replaced by the same solution, which now contained the DIG-labeled 1.3 kb laminin γ1 probe (200–400 ng/ml). The slides were heated for 5 min to 80°C. The sections were coverslipped with HybriSlip (Grace Bio-Labs) and placed in a humidified chamber in which Kimwipes were soaked in a solution containing 5× SSC and 50% formamide overnight at 72°C. The next day the coverslips were removed by placing the sections in 72°C, 5× SSC for 5 min. The slides were washed further in 0.2× SSC warmed to 72°C for 1.5 hr. The sections were rinsed in 0.2× SSC for 5 min at room temperature and incubated in buffer B1 (0.1 M Tris, pH 7.5, 0.15 M NaCl) for 5 min further at room temperature, followed by an 1 hr incubation at room temperature with 10% heat-inactivated goat serum in buffer B1. The sections were incubated in a humidified chamber at 4°C overnight in buffer B2 (1:5000; anti-DIG antibody in 1% heat-inactivated goat serum in buffer B1). The next day the hippocampal sections were rinsed three times in B1 buffer for 5 min on a shaker and equilibrated with B3 for 5 min, followed by an incubation with the developing buffer B4 (100 mg/ml nitroblue tetrazolium, 50 mg/ml 5-bromo-4-chloro-3-indolyl phosphate, 0.24 mg of levamisole) for 1.5 d at room temperature in the dark. The reaction was

<

(Figure legend continued.) γ1 chain (red). Note that the staining for laminin is situated between layers of neurocan staining. Scale bar, 100 μm. D, Higher magnification of the CA3 region of another hippocampal section showing the reduced staining of neurocan in the stratum lucidum (SL; marked by white dots) and stratum pyramidale (SP) and the increased staining for neurocan in the stratum oriens (SO). Scale bar, 100 μm (SR, stratum radiatum). E, Histological cross section through the hippocampus of a 1-year-old animal stained for the γ1 chain of laminin. Note that the pattern and extent of the pathway resemble that at P4. The cell bodies and apical dendrites of the pyramidal neurons in CA3 contain a filamentous network containing the γ1 chain of laminin. Additionally, a globular form of laminin is present in the cell body region but especially in the stratum lucidum. Scale bar, 100 μm. The positions from which F, G, and H were taken are shown in E. F, Higher magnification of laminin staining in the pyramidal cell bodies and proximal apical dendrites. Scale bar, 25 μm. G, Inner part of the stratum lucidum with its obvious globular form of the γ1 chain of laminin. Scale bar, 25 μm. The asterisk in E marks the end of the laminin pathway that appears to be at the CA3–CA2 border. A higher magnification of this border is shown in H. Scale bar, 25 μm.

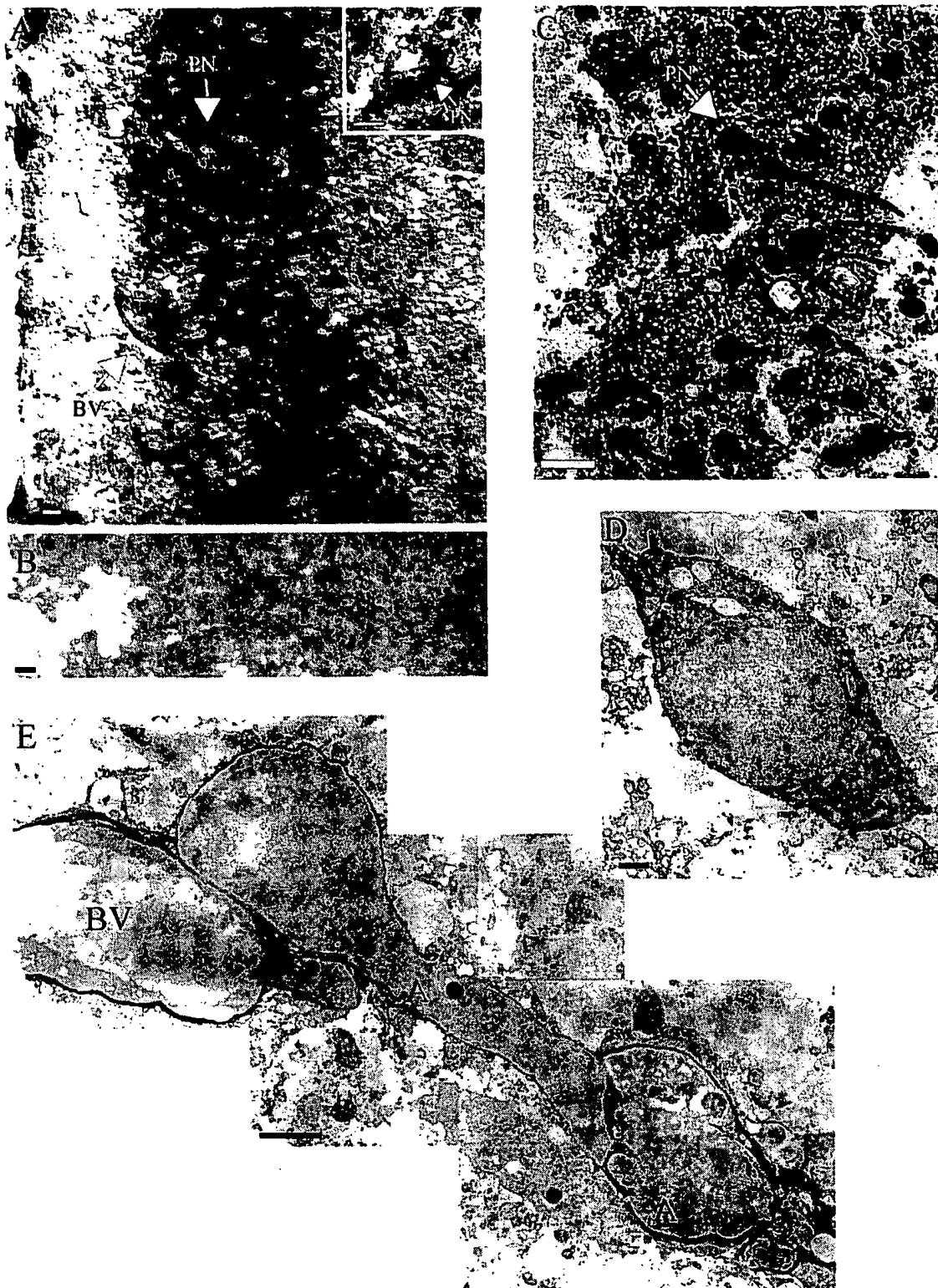


Figure 2. *A*, *In situ* hybridization against the $\gamma 1$ chain of laminin in the CA3 region of a P4 mouse hippocampus. The cell bodies of pyramidal neurons (*PN*) are stained (*BV*, blood vessel). The inset demonstrates a higher magnification confocal image of a labeled neuron (*PN*). Scale bars, 10 μm . *B*, CA1 region of the same section that lacks $\gamma 1$ chain mRNA. Scale bar, 10 μm . *C*, Semithin section (1 μm) of a P4 untreated/uncut rat hippocampal slice that had been in culture for 24 hr and stained for the laminin $\gamma 1$ chain. The cell bodies of pyramidal neurons (*arrow*) and their apical dendrites express laminin. Scale bar, 25 μm . *D*, *E*, EM immunohistochemical staining of the $\gamma 1$ laminin chain (no heavy metal counterstain was used) in ultrathin sections taken from the same slice from which the 1 μm section was taken (*BV*, blood vessel; *A*, astrocytes). Note in *D* the intracellular laminin reactivity within the cytoplasm of a pyramidal neuron cell body. Note also the intense extracellular staining associated with astrocyte membranes (*E*). Scale bar, 1 μm .

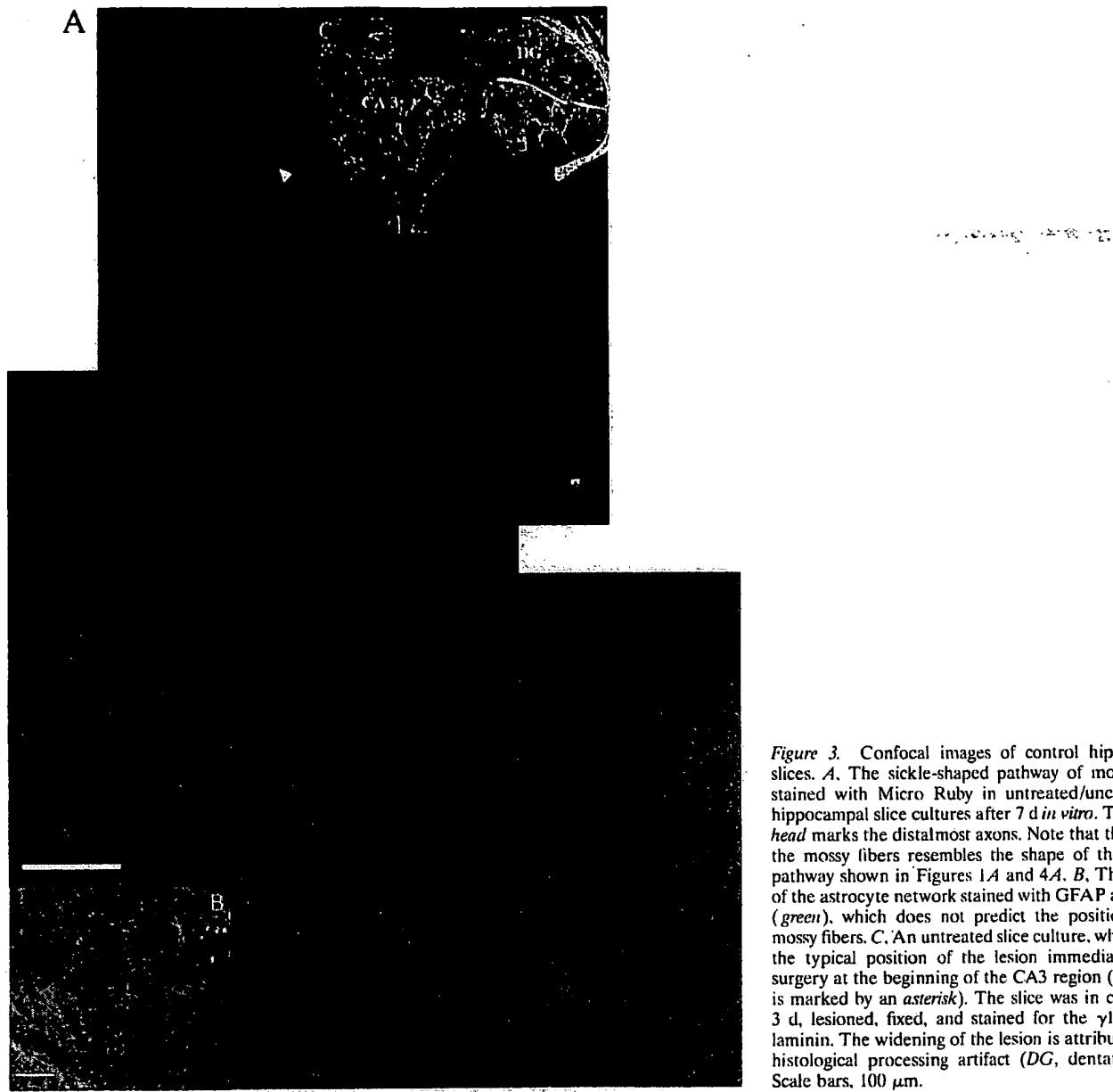


Figure 3. Confocal images of control hippocampal slices. *A*, The sickle-shaped pathway of mossy fibers stained with Micro Ruby in untreated/uncut rat P4 hippocampal slice cultures after 7 d *in vitro*. The arrowhead marks the distalmost axons. Note that the path of the mossy fibers resembles the shape of the laminin pathway shown in Figures 1*A* and 4*A*. *B*, The pattern of the astrocyte network stained with GFAP antibodies (green), which does not predict the position of the mossy fibers. *C*, An untreated slice culture, which shows the typical position of the lesion immediately after surgery at the beginning of the CA3 region (the lesion is marked by an asterisk). The slice was in culture for 3 d, lesioned, fixed, and stained for the $\gamma 1$ chain of laminin. The widening of the lesion is attributable to a histological processing artifact (*DG*, dentate gyrus). Scale bars, 100 μ m.

stopped by rinsing the sections in H₂O and mounting them with glycerol. The sections were viewed with a Leitz Orthoplan microscope (Wetzlar, Germany) under differential interference contrast optics and digitally photographed with an Optronics digital camera (Chelmsford, MA).

RNA isolation and PCR. After 7 d of treatment of three slice cultures each per concentration with antisense and mixed base ODNs (0.05 and 8 μ M), the slices were lysed and homogenized in Trizol (Invitrogen). The same procedure was performed on four slices after 3 d of treatment with 8 μ M DNA enzyme and control DNA enzyme (mixed base). Total RNA was prepared according to the supplier's protocol, adding yeast tRNA (Invitrogen) at a concentration of 10 μ g/ μ l as a carrier to help precipitation. From each sample 2 μ g (for the antisense group) or 3 μ g (for the DNA enzyme group) of total RNA was reverse transcribed into cDNA by using the GenAmp RNA PCR Core Kit (PerkinElmer Life Sciences and Roche, Emoryville and Palo Alto, CA) in accordance with the manufacturers' instructions. The step cycle program for the PCR was set to denaturation at 94°C for 30 sec, annealing at 65°C for 45 sec, and extension at 74°C for 40 sec for 30 cycles [laminin $\gamma 1$ -specific nested primers: rLamylas11, 5'-CCT CTC CGC CTC GTG GGC TT-3' and rLamyls11, 5'-CCC CTG TGG ACT CGG AGG CT-3' (Vanden Heuvel et al., 1996)]. This resulted in a product of 450 bp in length. β -Actin [β -actin-specific primers, acts 5'-ATC GTG GGC CGC CCT AGG CAC-3' and acts 5'-TGG CCT TAG-GGT TCA GAG GGG C-3' (Nudel et al., 1983)] were amplified as a control to test RNA integrity and to estimate the amount of RNA that was subjected to each PCR. The actin amplification was performed as described previously (Grimpe et al., 1999). The reaction products had a length of 240 bp. A PCR for the laminin $\beta 2$ chain was used for the DNA enzyme group as a control to show the specific digestion of the laminin $\gamma 1$ chain and the normal expression of other laminin chains. The conditions for the laminin $\beta 2$ chain PCR were the following: denaturing at 94°C for 30 sec, annealing at 67°C for 45 sec, and elongation at 74°C for 40 sec. The reaction formed a product of 349 bp. The sequences of the PCR primers were rLam β 2s, 5'-TCC AGA CCC CTA CAG CTC ACG G-3' and rLam β 2as, 5'-GCC CGT TGC ACT CAC ACT TCC G-3' (Hunter et al., 1989). All RT-PCRs were analyzed on 1.5% agarose gels, stained with

GAG ATC-3' (Vanden Heuvel et al., 1996). The step cycle program for the nested PCR was set to denaturation at 94°C for 30 sec, annealing at 65°C for 45 sec, and extension at 74°C for 40 sec for 30 cycles [laminin $\gamma 1$ -specific nested primers: rLamylas11, 5'-CCT CTC CGC CTC GTG GGC TT-3' and rLamyls11, 5'-CCC CTG TGG ACT CGG AGG CT-3' (Vanden Heuvel et al., 1996)]. This resulted in a product of 450 bp in length. β -Actin [β -actin-specific primers, acts 5'-ATC GTG GGC CGC CCT AGG CAC-3' and acts 5'-TGG CCT TAG-GGT TCA GAG GGG C-3' (Nudel et al., 1983)] were amplified as a control to test RNA integrity and to estimate the amount of RNA that was subjected to each PCR. The actin amplification was performed as described previously (Grimpe et al., 1999). The reaction products had a length of 240 bp. A PCR for the laminin $\beta 2$ chain was used for the DNA enzyme group as a control to show the specific digestion of the laminin $\gamma 1$ chain and the normal expression of other laminin chains. The conditions for the laminin $\beta 2$ chain PCR were the following: denaturing at 94°C for 30 sec, annealing at 67°C for 45 sec, and elongation at 74°C for 40 sec. The reaction formed a product of 349 bp. The sequences of the PCR primers were rLam β 2s, 5'-TCC AGA CCC CTA CAG CTC ACG G-3' and rLam β 2as, 5'-GCC CGT TGC ACT CAC ACT TCC G-3' (Hunter et al., 1989). All RT-PCRs were analyzed on 1.5% agarose gels, stained with

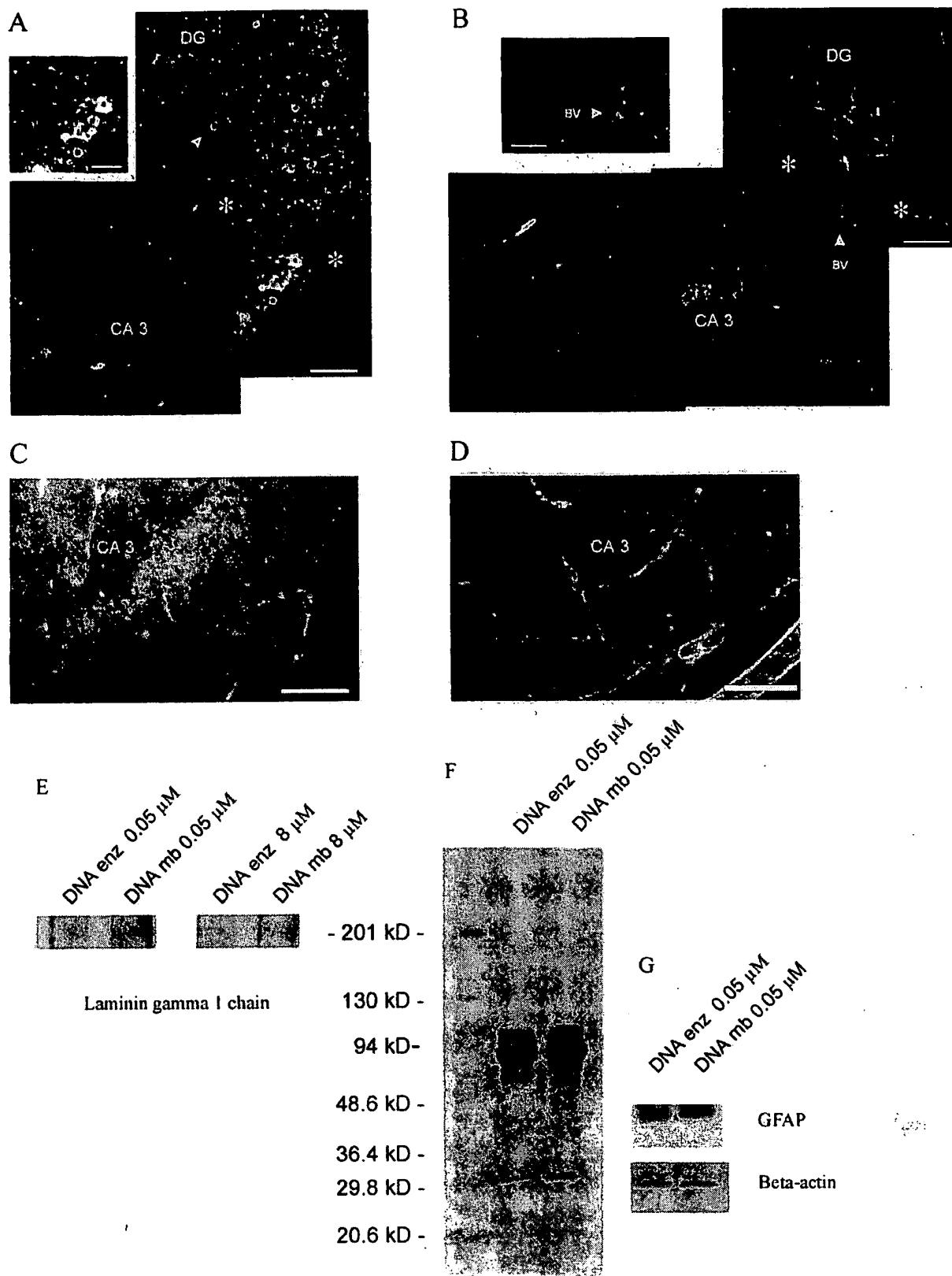


Figure 4. Laminin $\gamma 1$ staining of hippocampal slice cultures treated with antisense and mixed base ODNs as well as DNA enzyme and control DNA enzyme (**A–D**). Also shown is an immunoprecipitation of laminin $\gamma 1$ chain after DNA enzyme or control mixed base DNA enzyme (**E**). Coomassie staining (**F**) of protein extracts, and Western blots (**G**). **A**, **B**, Hippocampal slice cultures pretreated for 2 d with $0.1 \mu M$ mixed base ODNs (**A**) and with $0.1 \mu M$ antisense ODNs (**B**) against the $\gamma 1$ chain of laminin, transected (marked by asterisks), and treated for 5 d further (Figure legend continues.)

ethidium bromide, and photographed with a Polaroid camera (Kodak, Rochester, NY).

Southern blot analysis. The oligodeoxynucleotide *risLamylas1* [5'-CAG CAA GAG CCT TGG CAG CGT CGG CTC GAG CTA GGA GTT GGT CAG CGG TCT GCT G-3'] (Vanden Heuvel et al., 1996) obtained from a commercial supplier (MWG Biotech) specific for the laminin $\gamma 1$ chain was DIG labeled according to the manufacturer's protocol (Boehringer Mannheim). The amplified cDNA after antisense ODN or DNA enzyme treatment was separated electrophoretically on a 1.5% agarose gel. The gel was denatured for 15 min in 3 M NaCl and 0.4 M NaOH, followed by a 15 min incubation in transfer solution (3 M NaCl and 8 mM NaOH), and blotted onto Hybond N⁺ (Amersham Biosciences) as described previously (Sambrook et al., 1989). The next day the membrane was blocked and treated according to Engler-Blum et al. (1993). CDP-star (PerkinElmer Life Sciences and Roche) was used as a detection reagent. The blots were exposed to Amersham Hyperfilm for 1 and 2 sec.

RESULTS

In situ hybridization and immunohistochemical staining for laminin mRNA and protein in P4 and adult hippocampus and the relationship of mossy fibers to the laminin pathway in hippocampal slices

The laminin $\gamma 1$ chain is part of 10 of the 15 known laminins (for review, see Luckenbill-Edds, 1997; Libby et al., 1999). Thus we postulated that interfering with its synthesis would be maximally effective for the investigation of laminin function in our model. The known exceptions that do not express the $\gamma 1$ chain are laminin 5 (for review, see Luckenbill-Edds, 1997), which contains the $\gamma 2$ chain and is expressed in the basal membrane of blood vessels, and a newly discovered basement membrane-independent group of laminins (numbers 12–15; Libby et al., 1997), which contain a $\gamma 3$ chain (Koch et al., 1999).

In histological sections taken from P4 hippocampi of normal rats (Fig. 1*A,B*) a narrow, sickle-shaped pathway of diffuse, punctate, immunopositive $\gamma 1$ chain laminin staining was identified beginning in the hilus and extending along the CA3 pyramidal cell body layer (stratum pyramidale). Staining was also present in and around the portion of the apical dendrites closest to the cell body but was lacking entirely in the basal and distalmost portions of the apical dendrites of the young CA3 pyramidal neurons (see also Fig. 2*C*). Thus, laminin staining was confined strictly within stratum pyramidale and stratum lucidum, i.e., the identical terrain as the mossy fiber pathway (see O'Keefe and Nadel, 1978; Amaral and Dent, 1981) (see also the schematic in Fig. 6). Interestingly, this staining pattern ended abruptly at the CA3–CA2 border, a point beyond which the mossy fibers did not cross. A higher magnification of this pathway emphasized the diffuse but obvious staining of the pyramidal neuron cell bodies that extended into their apical dendrites (Fig. 1*B*).

A nonradioactive *in situ* hybridization from freshly prepared P4 mouse hippocampal sections clearly showed mRNA expression of

the $\gamma 1$ chain of laminin in the cell bodies of the CA3 pyramidal neurons (Fig. 2*A, inset*). Laminin mRNA expression was lacking in the CA1 region of the same sections (Fig. 2*B*) and was at background levels in the sense controls (data not shown). It is important to clarify that mouse hippocampus was chosen because of the availability of the mouse probe and because axonal plasticity in the hippocampus (Nieto-Sampedro and Bovolenta, 1990) as well as the patterning of laminin protein in the CA3 pyramidal neurons is like that of the rat (W. Halfter, personal communication).

Semithin section (light microscopy; Fig. 2*C*) and ultrathin section (EM; Fig. 2*D,E*) examination of immunopositive laminin $\gamma 1$ chain staining of P4 rat hippocampal slice cultures that had been *in vitro* overnight revealed three different structures that were associated closely with laminin. Under these conditions and fixation procedures these three were (1) the basal lamina surrounding blood vessels, (2) a dense network of ECM closely associated with the surface of astroglia and at contact points between astroglia in stratum lucidum (Fig. 2*E*), and (3) small deposits within the cytoplasm and at regions of close membrane contacts of CA3 pyramidal neurons (Fig. 2*D*). The pyramidal neuron staining visualized in semithin sections was, strikingly, confined to the cell body and clearly within the proximal portion of the apical dendrites in stratum lucidum (Fig. 2*C*).

These staining patterns, described here in finer detail, are in general agreement with those shown in the publications by Hagg et al. (1997) and Nakagami et al. [(2000), see their Fig. 2]. Such a laminin pathway was also detectable with the use of an antibody against the EHS form of laminin (laminin-1; data not shown). The pyramidal neurons also were stained positively (albeit weakly) with the $\gamma 1$ chain laminin antibody in freshly frozen, unfixed P4 hippocampi, ruling out possible confounding fixation artifacts. Control tissues were soaked in rabbit serum instead of the first antibody and/or visualized by staining with just the Alexa 594 (second antibody). In either case no staining beyond background was seen.

Laminin and neurocan form a pathway with boundaries

P4 hippocampi were stained simultaneously for laminin and for neurocan that, if positioned appropriately, could provide a potential inhibitory boundary on either side of the laminin pathway (Wilson and Snow, 2000). Strong staining for neurocan was present just peripheral to the pyramidal cell body layer (stratum oriens), a territory that overlaps the basal dendrites of the pyramidal neurons, and in stratum lacunosum (Fig. 1*C,D*). In general, neurocan staining was relatively lacking where laminin staining was present (Fig. 1*D*).

(Figure legend continued.) with the respective ODN concentrations. In the antisense-treated slice culture no laminin immunoreactivity was observed in the parenchyma (*B*), but some staining was present around the blood vessels (*BV*; arrow). However, in the mixed base-treated cultures (*A*) a sickle-shaped pathway of laminin staining can be observed in the pyramidal neurons and stratum lucidum, which foreshadows the mossy fiber pathway. Note the intensely stained cell bodies just distal to the lesion. Staining around the blood vessels (arrowheads) was present and can be used as an internal positive control for the antibody. Scale bar, 100 μ m (*DG*, dentate gyrus). Insets in *A* and *B* show higher magnifications of equivalent regions just distal to the lesion of the mixed base- and antisense ODN-treated slices. Scale bar, 50 μ m. *C*, The laminin $\gamma 1$ pathway in CA3 in an 0.1 μ M control DNA enzyme-treated slice that had been in culture for 4 d. Scale bar, 100 μ m. In comparison, *D* shows the same region in an 0.1 μ M DNA enzyme-treated slice. In the presence of control DNA enzyme the laminin $\gamma 1$ chain is expressed by the pyramidal neurons; however, such cells are not stained in the DNA enzyme slice. Blood vessel staining was still present. Scale bar, 100 μ m. *E*, An immunoprecipitation with a monoclonal antibody against the $\gamma 1$ chain of laminin in protein extracts from 12 slice cultures treated for 7 d with 0.05 μ M DNA enzyme (*DNA enz*), 0.05 μ M control DNA enzyme (*DNA mb*), 8 μ M DNA enzyme, and 8 μ M control DNA enzyme. The $\gamma 1$ chain has an expected size of 200 kDa. In the DNA enzyme-treated protein extracts this band is missing. The control DNA enzyme extracts still contained this band. *F*, The supernatant of the immunoprecipitation shown in *E* treated with 0.05 μ M DNA enzyme (*DNA enz*) and control DNA enzyme (*DNA mb*) stained with Coomassie blue in an SDS-PAGE gel. *G*, A Western blot for GFAP and β -actin from the same supernatant.

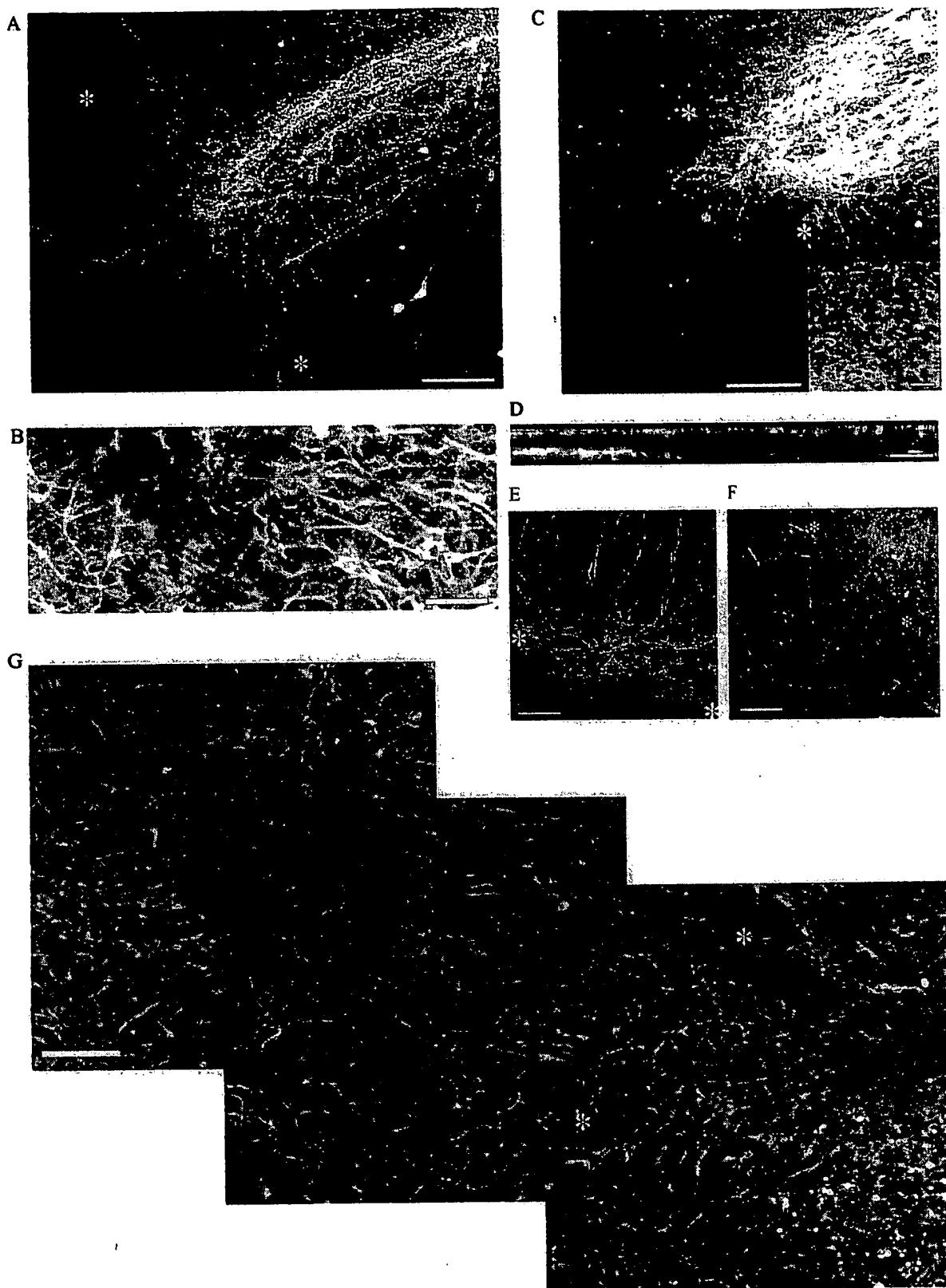


Figure 5. Hippocampal slice cultures treated with antisense ODNs, DNA enzymes, and mixed base ODNs. **A**, Slice culture pretreated with 8 μ M antisense ODNs for 2 d, lesioned (marked by asterisks), and stained 24 hr later with Micro Ruby. Note the abortive axon regeneration. Scale bar, 100 μ m. **B**, The GFAP staining of the astrocytes in the lesion site after 24 hr. Note that the astrocytes have refilled the lesion. Scale bar, 25 μ m. **C**, DNA enzyme-treated slice (8 μ M) that was labeled with Micro Ruby after 24 hr. Scale bar, 100 μ m. The inset shows the rapid (*Figure legend continues.*)

The laminin pattern in the adult hippocampus

In adult rat hippocampus the expression of the laminin $\gamma 1$ chain was seen distinctly as a system of fine filaments in the cell bodies and proximal portion only of the apical dendrites of the CA3 pyramidal neurons (Fig. 1E,F). The basal dendrites remained unstained. Additionally, in the pyramidal cell body layer, but especially in stratum lucidum, strong globular staining was associated with large numbers of small, irregularly shaped protrusions that emanated from the proximal apical dendrites (Fig. 1G). Such structures resembled dendritic spines and, again, their position overlapped precisely with that of the mossy fiber pathway (Chicurel and Harris, 1992; Harris and Kater, 1994) (see also the schematic in Fig. 6). In adult hippocampus the neurocan staining was reduced, which confirms a previous result of Haas et al. (1999). A higher magnification of the CA3–CA2 border from this adult hippocampus (Fig. 1H) shows the sudden decrease of laminin $\gamma 1$ chain expression at this location.

The development and lesioning of the mossy projection in slice cultures

Confocal microscope reconstructions of control P4 hippocampal slice cultures that were unlesioned, untreated, and allowed to develop *in situ* for 7 d further and then injected with Micro Ruby in the granule cell layer reveal the origin and entire sickle-shaped route of the mossy fibers (Fig. 3A). The trajectory of the axons, which passes through the astroglial framework of the slice (Fig. 3B), does not appear to be foreshadowed by the physical patterning of the glia, which are arranged in a contiguous meshwork that lacks linear organization. The mossy projection that begins in the dentate gyrus as a very tight bundle sometimes can spread out slightly in CA3 (Fig. 3A). The pattern of immunoreactive laminin $\gamma 1$ chain in hippocampal slices precisely mirrors the route of outgrowth of the mossy fibers (compare Figs. 3A, 4A).

Figure 3C is presented to give an overview of the position of the lesion that was used to sever the mossy axons. However, it should be noted that, in slice cultures that are stained with an antibody against the $\gamma 1$ chain of laminin immediately after the lesion, manipulation of the section without any time allowed for normal healing of the wound creates a gap between the cut edges of the tissue.

Immunohistochemical visualization and immunoprecipitation of laminin protein in hippocampal slices after antisense ODN and DNA enzyme treatment

In our experiments we treated hippocampal slice cultures with a variety of concentrations of antisense, mixed base (control) ODNs, DNA enzyme, and mixed base (control) DNA enzyme from 0.05 to 8 μM . These concentrations have been accepted as being within an optimal range by many other labs (Leslie et al., 1999). In all cases the effects on axon regeneration were mainly equivalent. In the description of the results that follows, representative data of selected concentrations are shown. However, for a complete analysis of the full range of the experiments, see Table 1 for quantitative data.

P4 hippocampal slice cultures treated with 0.1 μM antisense, 0.1

μM mixed based (control) ODN, 0.1 μM DNA enzyme, or 0.1 μM control (mixed base) DNA enzyme for 2 d were lesioned and maintained in culture for 1–5 d more in the continued presence of these reagents (Fig. 4A–D). The numbers of laminin $\gamma 1$ chain stained blood vessels had declined in all of these slices (i.e., control and antisense ODNs as well as DNA enzymes). This is likely attributable, in large part, to breakdown of the vasculature, which occurs normally within the slice. However, a few stained vessel basal laminae were still visible in all slices, which suggests that laminin within the basal lamina may be especially stable. In the control antisense and control DNA enzyme-treated slices, densely stained laminin-positive cells and their processes persisted in the pyramidal and lucidum layers for the length of the experiment (Fig. 4A,C, inset). The stained cells formed a sickle-shaped pathway that closely resembled that which was described in normal postnatal hippocampal tissues and that also resembles the route taken by normal as well as regenerating axons in the control slices (see below; compare Figs. 1, 4A). However, importantly, neither in the antisense ODN-treated slice cultures (Fig. 4B, inset) nor in the DNA enzyme-treated slices (Fig. 4D) was such a pathway of nonbasal lamina laminin detectable by immunohistochemistry.

To add further confidence to the immunohistochemical data, we performed an immunoprecipitation of the 200 kDa laminin $\gamma 1$ chain with the same antibody used for the immunostaining. In these studies we focused on and compared slices that were treated with DNA enzyme and control DNA enzyme. In these studies DNA enzyme or control DNA enzyme at a concentration of 0.05 or 8.0 μM was added to P4 slices for 7 d. The slices had been lesioned on day 2. Immunoprecipitation showed a lack of the 200 kDa band in the DNA enzyme-treated slices (Fig. 4E). In contrast, the control DNA enzyme-treated slices showed a strong positive band at 200 kDa (Fig. 4E). To make certain that the same amount of protein was loaded, we applied 10 μl of the same supernatant that was left over from the immunoprecipitation to an SDS-PAGE gel and stained it with Coomassie blue (Fig. 4F). Many comparably stained bands confirmed that equal amounts of total protein were present in each lane. To ascertain whether several selected proteins were still present that are unrelated to laminin, but for which the presence is indicative of the specificity of treatment as well as the overall health of the DNA enzyme-treated cultures, we showed that GFAP and β -actin were at comparable levels (Fig. 4G).

Inhibition of regeneration of the mossy fibers in DNA enzyme- and antisense-treated organotypic hippocampal slices

Slices that were pretreated for 2 d with mixed base (control) or antisense ODNs against the $\gamma 1$ chain of laminin then were lesioned and allowed to develop for as little as 1 or as long as 5 d more (i.e., up to a total of 7 d *in situ*). In the continued presence of control ODNs the mossy fiber pathway regenerated robustly. Figure 5G demonstrates the exceptional speed of regeneration in a control slice that had been treated with 8 μM mixed base ODN

(*Figure legend continued.*) infiltration of the lesion with astrocytes. Note the abortive axon regeneration. Scale bar, 50 μm . D, The cross-sectional (x-y) location of the mossy fibers shown in x-z orientation in A (mossy fibers, red; astrocytes, green). Arrowheads indicate the position of the lesion. Scale bar, 50 μm . E, A hippocampal slice culture treated with 2 μM antisense ODN for 5 d after lesion. Scale bar, 100 μm . F, A slice culture treated with 8 μM DNA enzyme for 5 d after lesion. Scale bar, 25 μm . G, A control hippocampal slice culture that was pretreated with 8 μM mixed base ODNs 2 d before being transected and allowed to regenerate for 1 d after the lesioning. The culture was injected with Micro Ruby (red) to label the regenerated mossy fibers and stained with GFAP antibodies to label the astrocytes (green). The lesion is marked by asterisks. Scale bar, 100 μm .

and allowed to regenerate for 24 hr. Indeed, the regrowing axons had the capacity to elongate approximately three-fourths of the way to the end of their pathway in just 1 d. Unlike untreated uncut slices (Fig. 3A), mixed base-treated lesioned slices showed a broadening of the fibers as they passed through the hilus as well as a slight contortion of the typical hook shape. It is noteworthy, however, that both of these variations, which likely are caused by misalignment of the stratum lucidum after the lesion, can be typical of hippocampal slices that are allowed to regenerate. The DNA enzyme controls (DNA mixed base, 8 μ M) also regenerated and were comparable with the antisense mixed base control slices (data not shown).

Treatment of P4 slices for 2 d with 8 μ M antisense ODN to the $\gamma 1$ chain of laminin, followed by transection and further treatment for 1 or 5 d additionally in culture (i.e., up to 7 d total), led to a dramatic diminution in the number of regenerating axons as well as a remarkable change in the anatomical pattern of axon regeneration that typically occurs in the lesioned slice (Fig. 5A,E). Importantly, regeneration was mainly abortive in slices treated with concentrations of this potent reagent as low as 0.05 μ M (see Table 1). Thus as early as 1 d (Fig. 5A) or as late as 5 d after lesion (Fig. 5E) the vast majority of the regenerating mossy fibers could not cross the injury site. They did regenerate, albeit slowly, within and along the lesion itself where the fibers appeared to fasciculate with each other (Fig. 5E). Axonal surfaces are an excellent substrate for regeneration, because they carry various growth-promoting adhesion molecules such as L1, neural cell adhesion molecule (NCAM), and N-cadherin (Walsh and Doherty, 1997; Woolhead et al., 1998). Importantly, a small but significant number of axons did navigate across the lesion successfully. However, once beyond the lesion these fibers also grew poorly in comparison to those in the lesioned mixed base-treated slices (compare Fig. 5G with A), and they took an uncontrolled and meandering course that was not at all confined to stratum lucidum or stratum pyramidale.

Treatment of lesioned hippocampal slices that used the 2 + 1 d treatment regimen with 8 μ M DNA enzyme caused a lack of regeneration that was essentially identical in pattern to that caused by antisense (Fig. 5C,F). Similar to the antisense experiments, in the presence of DNA enzyme directed against the $\gamma 1$ chain of laminin, the mossy fibers, although mainly unable to regenerate within stratum lucidum or stratum pyramidale, did continue to elongate, albeit slowly and haphazardly beyond, within, and on the proximal side of the lesion.

To provide a measure of confidence in regard to completeness of the transection in every slice, confocal scans along the x - y -axis (Fig. 5A) were rotated along the x - z -axis. As shown in Figure 5D, mossy fibers both proximal and distal to the lesion site were always in the middle of the slice [a typical appearance for the organotypic hippocampal slice culture (Buchs et al., 1993)] and never along the bottom where they would be expected to reside if the scalpel blade had pushed some fibers downward but had allowed for sparing.

Quantification of the amount and length of axon regeneration in antisense and mixed base-treated hippocampal slices

The results of treatment for 5 d after lesion (i.e., 2 + 5 d regimen) with five different concentrations of antisense ODNs (0.05 to 8 μ M) are shown in Table 1 and Figure 6. Measuring the lengths of individual mossy fibers is difficult because they are very thin and intertwined. Therefore, pixel intensity measurements of Micro

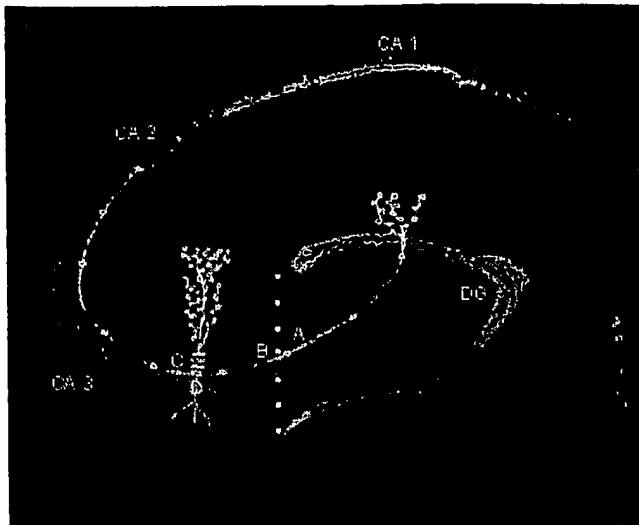


Figure 6. This figure is a composite of a schematic drawing adapted from O'Keefe and Nadel (1978) and a double immunohistochemical stain of a section of the hippocampus kindly reproduced with permission from Dr. Daniel Peterson and *Current Opinion in Pharmacology* [portion of a figure on the cover (2002)]. The green neuron cell bodies are stained with Neu N antibodies, and the mossy fibers among others have been stained red with calbindin antibodies. The schematic depicts an adult granule neuron in the dentate gyrus that extends a mossy fiber within the stratum lucidum (tight red bundle) to the border of CA3–CA2. A typical pyramidal neuron also is drawn to show that the mossy fiber projection within the stratum lucidum intersects only with a restricted portion of the proximal apical dendrite. Additionally, this picture shows the locations at which axonal densities were quantified. A is proximal to the lesion (measure point 1); B is distal to the lesion (measure point 2); C is 190–230 μ m distal to point B (measure point 3). Point A is in the dentate gyrus; points B and C are in the CA3 region. The lesion is shown by white dots.

Ruby-labeled axons from such slice cultures were made at three positions: (A) just proximal, (B) just distal, and (C) 190–230 μ m distal to lesion sites in the hilar portion of CA3 (Fig. 6). To standardize our measurements to account for interslice variability, we considered the proximal measurement in the dentate to be the 100% determination (see Table 1, position A). The two distal measurements (positions B and C in Table 1) were expressed as percentages of the proximal measurement for each slice. The furthest measurement from the lesion (position C), which was 190–230 μ m from the lesion, was chosen because it is a position in which significant numbers of axons can regenerate in all controls. The quantitation showed that, at position B just distal to the lesion, mixed base-treated slices had a higher regeneration rate (49–94%) than antisense-treated slices (20–35%). In addition, at the longer distance (position C) all mixed base-treated hippocampal slices showed a clear regeneration rate of 19–30%. However, in the antisense-treated cultures the vast majority of fibers had fallen considerably short of the greater distance; thus the quantitative measurements at point C were essentially at background levels. Considering all 30 slice cultures treated with all concentrations of antisense ODNs (Table 1), in only two of these 30 slices at one of the lowest concentrations (0.1 μ M) there was one lone fiber per slice that grew significantly. Thus in one treated slice a single mossy fiber reached the distance of 190 μ m, and in the other slice a single fiber reached nearly 250 μ m.

In the DNA enzyme-treated and control DNA enzyme-treated slices, axon lengths were not quantified. However, it was clear that the failure of regeneration brought about by the DNA enzyme

Table 1. Quantitation and statistical evaluation of regenerated fiber density in slice cultures treated with different concentrations of antisense and mixed base ODNs for 2 d, transected, and treated for 5 additional days

Mixed base concentration (μM)	Number of slices showing axon regeneration	Number of slices showing limited axon regeneration	Reg/SD point b $\times 100$ point a (%)	Reg/SD point c $\times 100$ point a (%)	Total number of slices
0.05	4	1	49.7/22.4	19.6/14.3	5
0.1	4	-	77.8/29.9	30.6/18.4	4
0.5	5	1	81.4/27.6	19.3/7.0	6
2	3	-	73.7/26.0	28.5/13.6	3
8	3	-	94.4/12.8	47.3/24.1	3

Antisense concentration (μM)	Number of slices showing axon growth inhibition	Number of slices showing some axon regeneration	Reg/SD point b $\times 100$ point a (%)	Reg/SD point c $\times 100$ point a (%)	Total number of slices
0.05	7	2	24.6/9.3	Bkg	9
0.1	8*	1	35.1/18.3	*	9
0.5	3	1	20.2/7.6	Bkg	4
2	5	-	28.8/11.0	Bkg	5
8	3	-	23.6/8.3	Bkg	3

Bkg, Background; Reg, regeneration. *In two slices from this group one mossy fiber per slice grew a relatively long distance (see Results).

was qualitatively equivalent to the antisense- and mixed base ODN-treated slice cultures (compare Fig. 5A with C).

Astroglial cells at the lesion site

Astrocytes completely repopulate the 50–100 μm wide gap created by the scalpel blade within the first day after injury (Fig. 5B,C, inset in C, G). Also, an aerial view of the overall shape of the astrocyte network (Fig. 5G) within the slice was neither predictive of where the laminin pathway was being expressed nor where the future route of mossy fiber regeneration would be (compare Figs. 4A, 5G). Importantly, in control slices the regenerating axons did not wander as they crossed the lesion site. Therefore, the lesion itself did not appear to be even a minimal barrier to the regenerating axons (Fig. 5G). Comparisons between the antisense- or DNA enzyme-treated slice cultures and their respective controls at all concentrations revealed that the shapes of the astrocytes throughout the slice and the rate at which astrocytes invaded the lesion gap were equivalent. Additionally, the levels of GFAP protein in DNA enzyme- and control DNA enzyme-treated slices were comparable when analyzed by Western blot analysis of SDS-PAGE (Fig. 4G).

Regrowth of the mossy fibers after termination and washout of antisense ODNs and DNA enzymes

This “recovery” experiment was undertaken to investigate further whether (1) the hippocampal slice cultures were in relatively good health and (2) whether they had the ability to regenerate (i.e., delayed regeneration) after the antisense ODNs or DNA enzymes were washed away.

Slice cultures of P4 rats were treated with 8 μM antisense ODNs or DNA enzyme for 2 d. Early in the morning of day 3 the slice cultures were lesioned and treated for 1 d more. On days 4–7 in culture, antisense or DNA enzyme treatment was stopped, and fresh medium was added. Thus, after a total of 7 d in culture the mossy fibers were labeled with Micro Ruby (Fig. 7A,B). In such antisense ODN- and DNA enzyme-treated/washout slice cultures the mossy fibers did, indeed, show a renewed burst of regeneration across the lesion site. Furthermore, the regrowing axons displayed a strong tendency to travel within strata lucidum and pyramidale. The lesion was not readily visible because astrocytes

invaded and refilled the lesion completely (Fig. 7C,E). Therefore, antisense or DNA enzyme treatment (at least for 3 d) did not appear to cause overt or irreversible harm to the mossy fibers, nor did the treatments appear to inflict damage to the glia or cause permanent damage to the pyramidal neurons that again were able to upregulate the production of laminin (Fig. 7D).

An analysis of the effects of antisense ODNs and DNA enzymes on mRNA levels

Neither of the concentrations of antisense ODNs (0.05 or 8 μM) produced a significant compensatory increase or decrease in laminin $\gamma 1$ chain mRNA expression when analyzed via RT-PCR/Southern blot techniques. Therefore, the antisense-treated slice cultures did not appear to upregulate mRNA production to compensate for the missing $\gamma 1$ chain of laminin (Fig. 8A). However, when the DNA enzyme-treated slices were analyzed in the same manner, specific digestion of the laminin $\gamma 1$ chain mRNA was seen (Fig. 8C). The specificity of the DNA enzyme reaction was shown by comparing a closely related mRNA for the laminin $\beta 2$ chain, which was unaltered (Fig. 8E). As a control for loading equal amounts of mRNA, β -actin mRNA was used in all cases (Fig. 8B,D).

DISCUSSION

In the present study we have focused on the question of whether a well described pathway of cells and extracellular matrix containing the $\gamma 1$ chain of laminin plays a critical role in support of CNS axon regeneration. To answer this question, we performed experiments with the use of cultured hippocampal slices because they retain much of the circuitry of the intact hippocampus and because *in situ* (as well as *in vivo*) the mossy fibers exhibit a profound capacity for compensatory sprouting (Altman and Das, 1965; Laurberg and Zimmer, 1981; Eckenhoff and Rakic, 1988; Eriksson et al., 1998; Mizuhashi et al., 2001) as well as regeneration (Zimmer and Gähwiler, 1987; Li et al., 1994; Nguyen et al., 1996; Eriksson et al., 1998). In the *in situ* model the mossy fibers can regenerate vigorously across a spontaneously repaired line of damage and along their proper pathway. This phenomenon may bear certain similarities to forms of reactive gliosis that are not

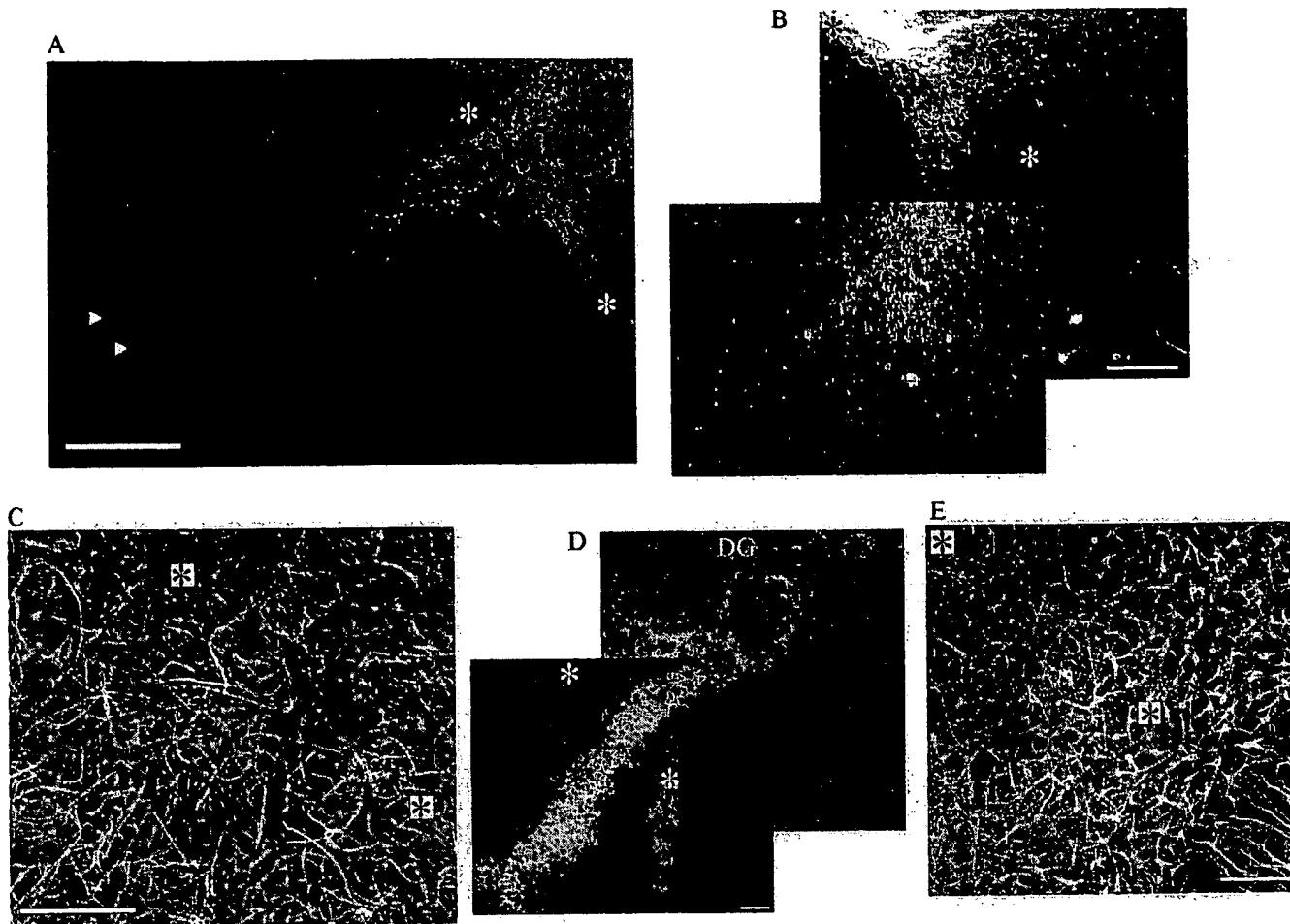


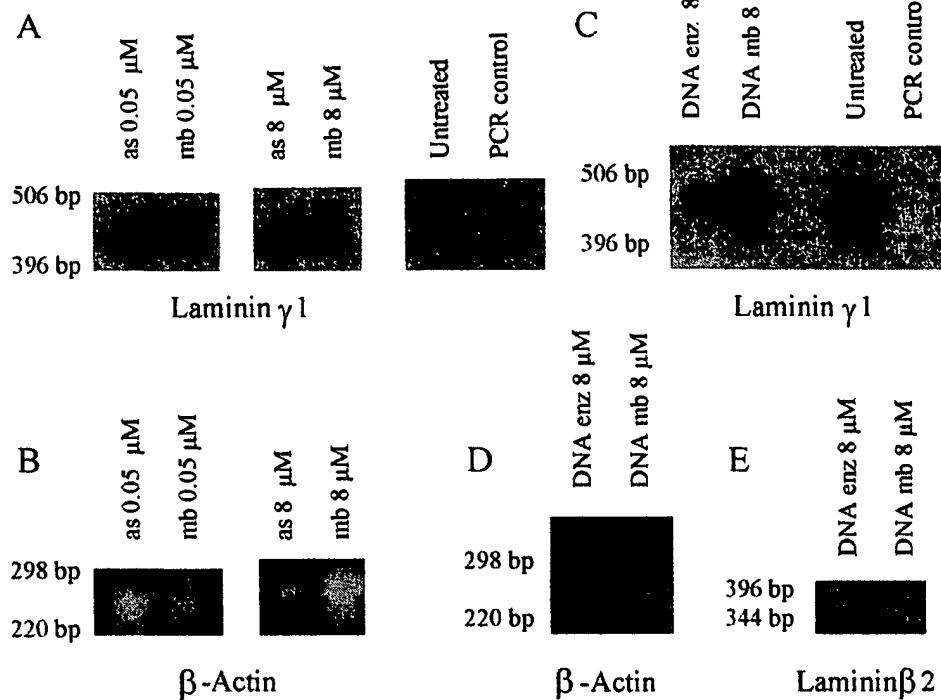
Figure 7. Recovery of the mossy fibers and reinitiated regeneration after washout of antisense ODNs and DNA enzyme in treated hippocampal slice cultures. *A*, A slice culture treated with 8 μ M antisense ODN for 3 d and then lesioned (marked by asterisks), followed by a washout of the antisense ODNs and no treatment for 5 d further. The mossy fibers regrow along their expected pathway. The ends of the mossy fibers are marked by arrowheads. *B*, Renewed axon regeneration in a slice culture treated with 8 μ M DNA enzyme for 3 d and lesioned (marked by asterisks), followed by a washout of the DNA enzyme and no treatment for 5 d further. *C*, GFAP staining around the lesion shown in *A*. The area of the lesion is marked with asterisks. *D*, The re-expression of the $\gamma 1$ chain of laminin in an 8 μ M antisense-treated slice culture. The slice had been treated for 3 d and lesioned (marked by asterisks), followed by a washout of the antisense ODN and no treatment for 5 d further (*DG*, dentate gyrus). *E*, The respective astrocyte staining of *B* around the lesion (marked by asterisks). Scale bars, 100 μ m.

associated with scar formation and allow for sprouting or regeneration *in vivo* (Altman and Das, 1965; Kaplan and Hinds, 1977; Raisman, 1985; Smith et al., 1986; Chauvet et al., 1995; Kuhn et al., 1996). In the laminin antisense ODN- and DNA enzyme-treated hippocampal slice cultures, although astroglial healing of the lesion site as well as overall glial patterning throughout the slice were unaffected, regrowth of the vast majority of mossy fibers was disallowed.

Thus, our data demonstrate for the first time a critical role for laminin(s) in an early postnatal regeneration model of the CNS that takes place in the context of a complicated array of interacting glial and neuronal cell types. Laminin that is embedded in the basal lamina of blood vessels is unlikely to play a role in axon regeneration in this model because its lacks appropriate spatial prepatternning and because vessel basal lamina is enwrapped by other molecules such as proteoglycans and collagens that are mainly nonpermissive to axon outgrowth (Burg et al., 1996; Joosten et al., 2000). The two additional forms of laminin (astrocyte and neuron-associated) within this specialized portion of the

CNS that could play a more obvious role in regeneration appear to be present in several different patterns and in tissue compartments both inside and outside of cells. Whether one or both of these basal lamina-independent types is responsible for axon regeneration is a matter of speculation, because both were reduced or eliminated during treatment. Therefore, we still do not know for certain which cell type or types express the axon regeneration-promoting form of laminin, nor do we know which form of laminin permits such robust regeneration to occur. Nonetheless, the precise spatiotemporal correlation that exists between regenerating granule cell mossy fibers and restricted portions of the CA3 pyramidal neurons that contain laminin protein as well as laminin $\gamma 1$ chain mRNA does lead us to suggest that a causal relationship mediated by laminin must exist between these two neuronal partners.

Laminin deposits along a filamentous network within the cell bodies and continuing into a restricted portion of the apical dendrites of CA3 pyramidal neurons (Fig. 2C) have not been described previously, although Hagg et al. (1989) did suggest that



349 bp). This related chain to the $\gamma 1$ chain of laminin is not affected by the DNA enzyme treatment. The PCR control is an RT-PCR without RNA but with all buffers to check for contamination.

hippocampal laminin was present at synapses. The role of laminin in the development and function of synapses on hippocampal pyramidal neurons is now under investigation by other labs where it has been shown that laminin may be involved with the regulation of long-term potentiation (Nakagami et al., 2000). It also is known that laminin has a strong organizing effect on the formation of point contacts and integrin clustering, which occur along the surfaces of axonal growth cone filopodia (Schmidt et al., 1995; Renaudin et al., 1999). In addition, laminin has been shown to have prominent enhancing effects on the motility of dendritic spines (Seil, 1998) for which the dynamic interactions with axonal growth cone filopodia may be essential during axonal outgrowth and synapse formation (Tsui et al., 1985; Halfter, 1996; Wu et al., 1999; Jontes and Smith, 2000; Korkotian and Segal, 2001; Prange and Murphy, 2001). It may be that pyramidal cell laminin, if transported and/or secreted locally within the vicinity of developing proximal spines [i.e., the so-called thorny excrescences, Gonzales et al. (2001) and Amaral and Dent (1981), see their Fig. 5], may play a critical role in stimulating exploratory behaviors by fine caliber dendritic protrusions for which the role may be to lure mossy fiber growth cone filopodia toward their surfaces. It is fascinating to consider the possibility that the highly restricted pathway of laminin on the apical dendrites of CA3 pyramidal neurons would place a potent axon regeneration-promoting molecule at the very doorstep of potential synaptic sites between regenerating mossy fiber terminals and their specific postsynaptic partners, a remarkably pinpointed distribution, indeed.

Laminin can be localized to focal contacts between glia and other non-neuronal cells or more diffusely in the extracellular matrix where it is represented by an immunoreactivity that is distributed in fine punctate deposits (Liesi and Silver, 1988). It has been demonstrated repeatedly that extracellular astroglial

Figure 8. RT-PCR analyses from RNA preparations taken from untreated slices as well as from slices treated with antisense (as), mixed base ODNs (mb), DNA enzyme (DNA enz), and mixed base DNA enzyme (DNA mb). The slices (treated and untreated) were maintained in culture for 7 d for antisense and mixed base ODNs and for 4 d for DNA enzyme and control DNA enzyme. **A**, RT-PCR Southern blot for the $\gamma 1$ chain of laminin after antisense and mixed base ODN treatment (expected RT-PCR product size, 450 bp). **B**, Agarose gel of an RT-PCR for β -actin mRNA from the same preparations shown in **A** (expected RT-PCR product size, 260 bp). Note that the laminin $\gamma 1$ chain mRNA shows only minimal changes, which are repeated in the size of the actin bands. **C**, RT-PCR Southern blot for the $\gamma 1$ chain of laminin from slice cultures treated with DNA enzyme showing a reduction of $\gamma 1$ chain mRNA compared with slices treated with control DNA enzyme (expected RT-PCR product size, 450 bp). **D**, Agarose gel of an RT-PCR for β -actin mRNA from the same preparation as shown in **C** (expected RT-PCR product size, 260 bp). **E**, The use of the same preparation as in **C** for an RT-PCR of the laminin $\beta 2$ chain (expected RT-PCR product size,

matrices, *in vitro*, are axon growth-supportive, at least in part, because of laminin (Manthorpe et al., 1983; Liesi and Risteli, 1989; Smith et al., 1990; Wujek et al., 1990; Groves et al., 1993; Tomasselli et al., 1993; Garcia-Abreu et al., 1995). However, the presence and/or function of this diffuse form of laminin, *in vivo*, have been extremely controversial. Indeed, only a few groups of researchers have been able to visualize the extracellular punctate form of laminin within the brain parenchyma (Letourneau et al., 1988; Liesi and Silver, 1988; McLoon et al., 1988; Gordon-Weeks et al., 1989; Liesi and Risteli, 1989; Zhou, 1990; Cohen and Johnson, 1991). Now a number of labs clearly have revealed a diffuse, punctate laminin immunoreactivity in the stratum lucidum and area CA3 of the P4 hippocampus (Hagg et al., 1989, 1997; Zhou, 1990; Nakagami et al., 2000; this study). The extracellular location of laminin, which was confirmed here by EM, suggests the possibility that axon guidance information may be generated by the expression of certain forms of this family of ECM molecules that develop within confined macroscopic highways.

Abundant quantities of laminin in the extracellular space of stratum lucidum not only may stimulate regenerating axons as they navigate through this region but also may be involved with interactions between the glia and/or between glia and neurons that are critical in the assembly or maintenance of the three-dimensional geometry of the glial/neuronal scaffold (Willbold et al., 2000). Perhaps laminin is involved with specialized astroglial movements (e.g., axon-enwrapping or channel-forming behaviors) that are critical as axonal growth cones pass over their membranes (Smith et al., 1986; Payne and Lemmon, 1993; Isaacson et al., 1995). Such movements may help to allow for some form of structural plasticity that occurs amid a denser and more rigid cellular backbone of tissue. Although it is clear that laminin antisense ODN and DNA enzyme treatment did spare gross glial

architecture while severely altering axon regeneration, we cannot rule out the possibility that subtle glial movements in association with axonal regeneration were altered.

Is laminin the only regeneration-promoting molecule in the hippocampus?

Although reduction of laminin expression by using antisense ODN and DNA enzyme technology curtails the vast majority of regenerating mossy fibers, a small number of mossy fibers can grow across the lesion, at least for short distances. The simplest explanation for the incomplete effect on regeneration is that antisense ODNs and even DNA enzymes do not eliminate 100% of laminin expression or that laminin degradation is not fully complete at the time these fibers cross the lesion. Another possibility is that reduction of $\gamma 1$ subunit expression produces an upregulation of another laminin subunit (i.e., other than $\beta 2$, such as $\alpha 5$ and $\gamma 3$, which have also been identified in the CNS; W. Brunkent, personal communication) that compensates for the loss of $\gamma 1$ (Libby et al., 1997, 1999; Patton et al., 1997). Also, although we have manipulated specifically the expression of the $\gamma 1$ chain of laminin, it would be premature to claim that the effects on axon outgrowth are mediated solely or directly by laminin. For example, the loss of laminin could affect indirectly the function of other molecules. Laminin is known to bind to heparan sulfate proteoglycans such as syndecan, agrin, and perlecan, which, in turn, can bind a variety of growth factors and neurotropic molecules (Olsen, 1999). Future experiments in which integrin expression in granule cell neurons is reduced by using DNA enzymes and antisense ODNs will help to clarify the possibility of a direct role for laminin in CNS axon outgrowth and regeneration. In addition to laminin, a number of other axon guidance molecules, including netrin-1 (Mitchell et al., 1996; Steup et al., 2000), polysialylated NCAM (Seki and Arai, 1991; Seki and Rutishauser, 1998; Cremer et al., 2000), and L1 (Miller et al., 1993; Wong et al., 1995), are expressed in the hippocampus and could affect mossy fiber regeneration. However, they do not appear to be capable of compensating in a major way for the lost members (either bound or soluble forms) of the laminin family during regeneration.

Why does the hippocampus maintain a pathway that expresses such high concentrations of a potently growth-supporting form of laminin?

Evidence exists that small populations of neurons continue to be born, fully mature, and to elaborate axons (van Praag et al., 2000) in the adult hippocampus (Eckenhoff and Rakic, 1988; Eriksson et al., 1998). The newly born neurons originate from putative stem cells that exist in the subgranular zone of the dentate gyrus (Eriksson et al., 1998; Seri et al., 2001). Progeny of these immature cells differentiate into neurons in the granule cell layer within a month of their birth. This late form of neurogenesis and axon growth continues throughout the adult life of the rodent (Gage et al., 1995) as well as humans (Eriksson et al., 1998). The critical question arises: Does the laminin pathway in the hippocampus continue to exist to provide for *de novo* formed hippocampal neurons their differentiation signals as well as a guidance highway for their axons? Transplantation experiments that use stem cells precisely delivered within the territory of the normal or perturbed laminin pathway will help to address this question.

In conclusion, our experiments have demonstrated a critical role for laminin(s) in directing mossy fiber regeneration. Given the potent effects of laminin on promoting the outgrowth of CNS axons in the regenerating hippocampus, it is possible that the particular format in which laminin is found in this specialized

region of the brain also may be used in a strategy for stimulating the regeneration of other severed axons, such as those in the spinal cord, which fail to regenerate after injury.

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